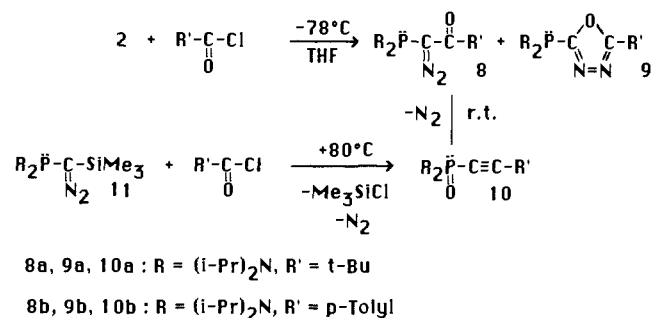
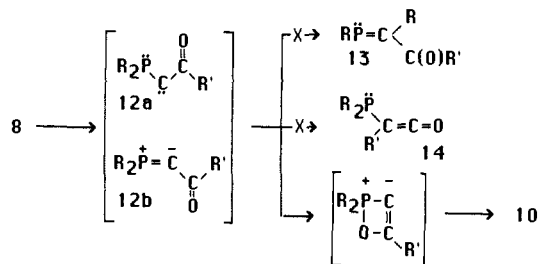


Scheme II



Scheme III



row element is involved, the carbenoid character is competitive, as recently shown for -S-N,⁸ -C-SF₃,⁹ and even -Si-Si-¹⁰ derivatives.

Acknowledgment. We thank the CNRS (GRECO Basses Coordinences) for support of this research.

Supplementary Material Available: Microanalytical, mass spectral, IR, and NMR (¹H, ¹³C, ³¹P, ¹⁵N) data (3 pages). Ordering information is given on any current masthead page.

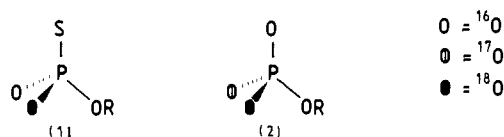
- (7) Curtis, T. *Ber. Dtsch. Chem. Ges.* **1889**, 22, 2161.
(8) Glemser, O.; Mews, A. *Angew. Chem., Int. Ed. Engl.* **1980**, 19, 883 and references cited therein. Atkinson, R. S.; Judkins, B. D. *J. Chem. Soc., Perkin Trans.* **1981**, 2615.
(9) Pötter, B.; Seppelt, K.; Simon, A.; Peters, E. M.; Hettich, B. *J. Am. Chem. Soc.* **1985**, 107, 980.
(10) Sekiguchi, A.; Zigler, S. S.; West, R., unpublished results.

Thiophosphoryl-Transfer Reactions: A General Synthesis and Configurational Analysis of O-Substituted [¹⁶O,¹⁸O]Thiophosphates

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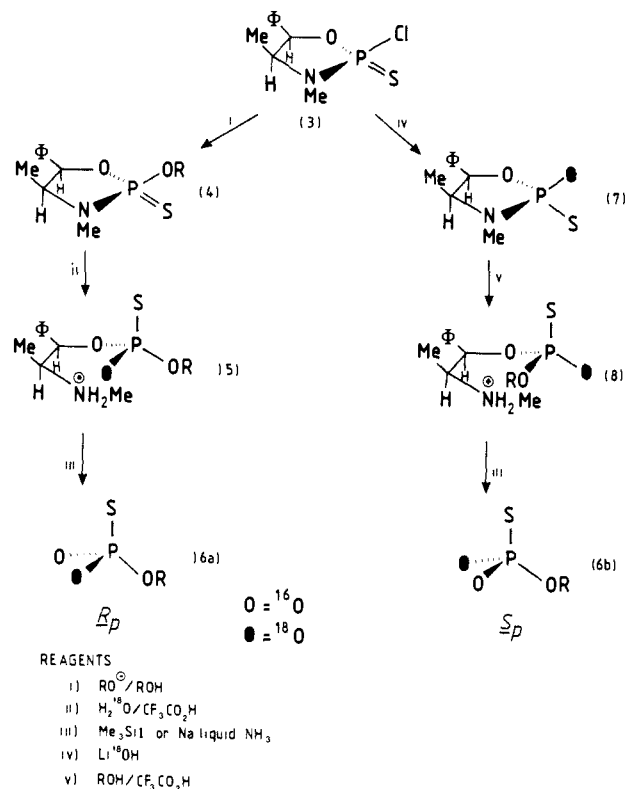
[¹⁶O,¹⁸O]Thiophosphate (1) and [¹⁶O,¹⁷O,¹⁸O]phosphate (2) esters have been utilized extensively to determine the stereochemical course of many enzyme-catalyzed thiophosphoryl-¹ and



phosphoryl-transfer² reactions. Although the stereochemical

- (1) Eckstein, F. *Angew. Chem., Int. Ed. Engl.* **1983**, 22, 423. Frey, P. A. *Tetrahedron* **1982**, 38, 1541.
(2) Knowles, J. R. *Annu. Rev. Biochem.* **1980**, 49, 877. Lowe, G. *Acc. Chem. Res.* **1983**, 16, 244. Gerlt, J. A.; Coderre, J. A.; Mehdi, S. *Adv. Enzymol.* **1983**, 55, 291.

Scheme I



courses of some simple chemical phosphoryl-transfer reactions have recently been determined,^{3,4} hitherto simple thiophosphoryl-transfer reactions have not been studied. With existing methods these would in fact be difficult to determine. Such studies would be of interest since (i) the stereochemical course of enzyme-catalyzed thiophosphoryl-transfer reactions has frequently been assumed to be the same as for the natural phosphoryl-transfer reaction and it would be pertinent to determine whether these reactions are indeed stereochemically equivalent⁵ and (ii) thiophosphate monoesters have been reported to react more rapidly via a dissociative reaction than the corresponding phosphate esters.⁶ We report here the first simple chemical configurational analysis of structures such as 1 (R = alkyl or aryl)⁷ together with general synthetic routes to simple [¹⁶O,¹⁸O]thiophosphate monoesters (1).⁸

Our two general routes to isotopically chiral [¹⁶O,¹⁸O] (or [¹⁷O])thiophosphate monoesters of either the R_p or S_p absolute configuration are shown in Scheme I. By analogy with the previously published route(s) to [¹⁶O,¹⁷O,¹⁸O]phosphate esters,⁹

- (3) Buchwald, S. L.; Knowles, J. R. *J. Am. Chem. Soc.* **1982**, 104, 1438. Buchwald, S. L.; Friedman, J. M.; Knowles, J. R. *J. Am. Chem. Soc.* **1984**, 106, 4911. Friedman, J. M.; Knowles, J. R. *J. Am. Chem. Soc.* **1985**, 107, 6126.
(4) Cullis, P. M.; Rous, A. J. *J. Am. Chem. Soc.* **1985**, 107, 6721. Cullis, P. M.; Rous, A. J. *J. Am. Chem. Soc.* **1986**, 108, 1298.
(5) The demonstration for a number of enzymes that phosphoryl and thiophosphoryl transfer proceed with the same stereochemical course (see ref 1 and 2) would suggest that within the constraints of the enzyme active site these two reactions are equivalent.
(6) Breslow, R.; Katz, I. *J. Am. Chem. Soc.* **1968**, 90, 7376.
(7) Two configurational analyses have been reported for AMPS-¹⁸O and other nucleoside [¹⁸O]thiophosphates: the first relies on the stereospecific enzyme-catalyzed phosphorylation of the pro-R/S oxygen as the key step (Sheu, K.-F. R.; Frey, P. A. *J. Biol. Chem.* **1977**, 252, 4445); the second method has assigned the absolute configurations of the O,S-dimethyl nucleoside triesters by relating these to the O-methyl nucleoside diesters which have been assigned on the basis of the known stereoselectivity of snake venom phosphodiesterase (Cummins, J. H.; Potter, B. V. L. *J. Chem. Soc., Chem. Commun.* **1985**, 851). Neither method was suitable for our proposed study.
(8) Previous syntheses of isotopically chiral thiophosphate monoesters based on the meso-hydrobenzoin route (Cullis, P. M.; Lowe, G. *J. Chem. Soc., Perkin Trans. 1* **1981**, 2317. Jarvest, R. L.; Lowe, G. *J. Chem. Soc., Chem. Commun.* **1979**, 364) have been reported but not extensively applied. Similarly [^{γ-16}O,¹⁸O,S]ATP and [¹⁸O]AMPS have been synthesized by routes that would not easily be extendible to simple thiophosphate esters.

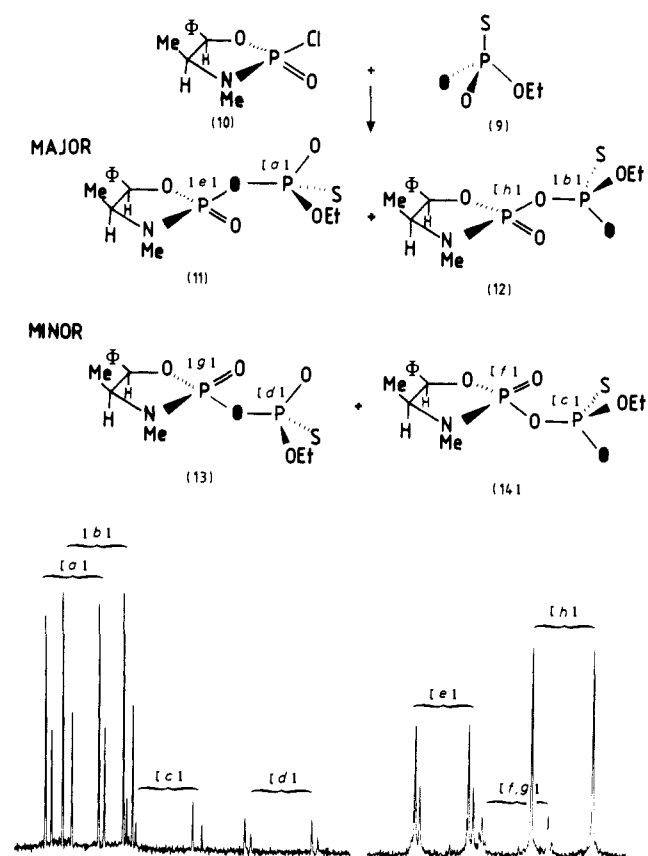


Figure 1. Stereochemical analysis of ethyl (S_p)-[^{16}O , ^{18}O]thiophosphate by ^{31}P NMR spectroscopy of the product following reaction with *cis*-2-chloro-3,4(*S*)-dimethyl-5(*S*)-phenyl-1,3,2-oxazaphospholidin-2-one. The spectrum was recorded on a Bruker AM-300 at 121.5 MHz and processed with Gaussian multiplication (Gaussian broadening 0.1 Hz, line broadening -0.3 Hz). The assignments are as shown with the downfield resonances (thiophosphoryl center) at ca. +46 ppm and the upfield resonance (1,3,2-oxazaphospholidine center) at ca. +7 ppm.¹²

these syntheses exploit the stereocontrolled displacement reactions of 2-substituted 1,3,2-oxazaphospholidine-2-thiones, which have established precedent in the work of Inch et al.¹⁰

The major objective has been the development of a general method for the configurational analysis of isotopically chiral thiophosphate monoesters. During the course of our work on the stereochemistry of phosphoryl transfer from P^1, P^1 -disubstituted pyrophosphates,⁴ we synthesized the unlabeled diastereomeric pyrophosphates corresponding to **11** and **12**. These were readily distinguished by high-field ^{31}P NMR spectroscopy and form the basis of the configurational analysis reported here. S_p -*O*-Ethyl [^{16}O , ^{18}O]thiophosphate (**9**) (^{18}O enrichment ca. 33%) was synthesized by the route shown in Scheme I. The absolute configuration follows from the synthesis. Reaction of **9** with the *cis*-2-chloro-1,3,2-oxazaphospholidin-2-one (**10**) derived from (-)-ephedrine gave rise to the pyrophosphate derivatives **11**–**14**. The high-field ^{31}P NMR spectrum together with the assignments are shown in Figure 1. Resonances corresponding to centers **e** and **h** can be unambiguously assigned since the 1,3,2-oxazaphospholidine phosphorus center is attached to ^{18}O in diastereoisomer **11** but not in diastereoisomer **12**, hence only one set of resonances will be split by the stereospecific incorporation of ca. 33% ^{18}O . On the basis of the bond-order dependence of the ^{18}O shift¹¹ on

(9) Abbott, S. J.; Jones, S. R.; Weinman, S. A.; Knowles, J. R. *J. Am. Chem. Soc.* **1978**, *100*, 2558. The extension of the published route to isotopically chiral phosphate monoesters to the synthesis of thiophosphate monoesters is nontrivial; the thiophosphorochloridate is significantly less reactive, the acid ring opening step can lead to competing loss of sulfur, and finally the removal of the ephedrine framework is difficult. Full details will be published elsewhere.

(10) Cooper, D. B.; Hall, C. R.; Harrison, J. M.; Inch, T. D. *J. Chem. Soc., Perkin Trans. 1* **1977**, 1969.

the thiophosphoryl signal, resonances **a** can be assigned to the diastereoisomer **11** in which the ^{18}O is located in the bridging position and resonances **b** can be assigned to the diastereoisomer **12**. The additional minor resonances seen in Figure 1 are due to structures **13** and **14**, which are epimeric at the ring phosphoryl center with respect to **11** and **12**.¹³ R_p -*O*-Ethyl [^{16}O , ^{18}O]thiophosphate would give rise to a ^{18}O shift on **h** rather than **e** and the magnitude of the ^{18}O shifts on **a** and **b** would be reversed. The downfield ^{31}P resonances arise from diastereoisomer **11** (in which the new chiral center at the thiophosphoryl position has the R_p configuration) while the upfield ^{31}P NMR resonances arise from diastereoisomer **12** (in which the new chiral center has the S_p configuration). We have established that this assignment holds for **6a** ($R = p$ -nitrophenyl) and **6b** ($R = \text{ethyl}$), and it may hold for a wide range of R groups. The above assignments form the basis of our method for studying the stereochemical course of simple thiophosphoryl-transfer reactions.¹⁴ The above analysis strategy is potentially general and may allow extension to the study of a range of hydrolysis reactions leading to phosphorus acids of the type $\text{R}^1\text{R}^2\text{PO}_2^-$ ($\text{R}^1 \neq \text{R}^2$).¹⁵

Acknowledgment. This work was supported by a grant from the SERC.

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(12) The ^{31}P NMR data from the spectrum shown in Figure 1 are as follows: diastereoisomer **11** δ (CDCl_3) +7.14 (d, $J_{\text{PP}} = 25.9$ Hz, 1,3,2-oxazaphospholidin-2-one, ^{18}O shift 2.28 Hz), +46.39 (d, $J_{\text{PP}} = 25.9$ Hz, R_p thiophosphoryl center, ^{18}O shift 2.84 Hz); diastereoisomer **12** δ (CDCl_3) +6.65 (d, $J_{\text{PP}} = 29.7$ Hz, 1,3,2-oxazaphospholidin-2-one), +46.29 (d, $J_{\text{PP}} = 29.7$ Hz, S_p thiophosphoryl center, ^{18}O shift 4.46 Hz).

(13) The trans diastereoisomers **13** and **14** apparently do not arise from trace amounts of the trans chloro compound analogous to **10** but are due to an epimerization reaction.

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Thiophosphoryl-Transfer Reactions: Stereochemical Course of Solvolysis of *p*-Nitrophenyl Thiophosphate in Protic Solvent and the Possible Role of Thiometaphosphate

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There is much current interest in monomeric metaphosphate as a possible intermediate in nucleophilic displacement reactions of monosubstituted phosphate esters¹ and, in particular, in relation to enzyme-catalyzed phosphoryl-transfer reactions.² Stereochemical,^{3,4} kinetic,^{5,6} and thermodynamic⁷ evidence suggests that metaphosphate is so reactive that it does not have a significant lifetime in *protic* solvents (although many other three-coordinate P(V) compounds have appreciable stabilities^{8,9}). In contrast,

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(2) Knowles, J. R. *Annu. Rev. Biochem.* **1980**, *49*, 877. Lowe, G. *Acc. Chem. Res.* **1983**, *16*, 244.
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(5) Skoogs, M. T.; Jencks, W. P. *J. Am. Chem. Soc.* **1983**, *105*, 3356. Skoogs, M. T.; Jencks, W. P. *J. Am. Chem. Soc.* **1984**, *106*, 7597.
(6) Bourne, N.; Williams, A. *J. Am. Chem. Soc.* **1983**, *105*, 3357. Bourne, N.; Williams, A. *J. Am. Chem. Soc.* **1984**, *106*, 7591.
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(8) Regitz, M.; Maas, G. *Top. Curr. Chem.* **1981**, *97*, 71–120.
(9) Roesky, H. W.; Ahlrichs, R.; Brode, S. *Angew. Chem., Int. Ed., Engl.* **1986**, *25*, 82.